

## Modeling the sodium conductance

Selina Baeza-Loya

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### Introduction

To clear up confusion and mollify any misunderstandings, I have typed up the modeling of sodium conductance using Hodgkin-Huxley (HH) formulations, as I understand them. I've included some inserts from the original HH papers (specifically, the 1952d paper, A QUANTITATIVE DESCRIPTION OF MEMBRANE CURRENT AND ITS APPLICATION TO CONDUCTION AND EXCITATION IN NERVE), as well as some other works that I've referenced over the years.

### Sodium conductance modeled as a set of first order equations

To model a traditional, TTX-sensitive transient sodium current, as was recorded in the giant squid axon by Hodgkin and Huxley, we begin with:

$$g_{Na} = \bar{g}_{Na} m^3 h$$

where  $g_{Na}$  is the sodium conductance,  $\bar{g}_{Na}$  is the fixed sodium conductance density of open Na channels,  $m$  is the activation rate and  $h$  is the inactivation rate.

Hodgkin and Huxley described the activation and inactivation with the following gating equations:

$$m' = \alpha_m(1 - m) - \beta_m m$$

$$h' = \alpha_h(1 - h) - \beta_h h$$

where  $\alpha$  and  $\beta$  are rate constants. See page 512, Hodgkin and Huxley 1952d.

To characterize voltage-gated channels, equations are fit to voltage clamp data. Under voltage clamp conditions, where the voltage can be held constant, the nonlinear gating equations can be reduced to:

$$m = m_{\infty} - (m_{\infty} - m_0) \exp\left(-\frac{t}{\tau_m}\right); h = h_{\infty} - (h_{\infty} - h_0) \exp\left(-\frac{t}{\tau_h}\right)$$

Where  $m_{\infty}$  is the steady-state activation function,  $\tau_m$  is the time constant of activation,  $h_{\infty}$  is the steady-state inactivation function, and  $\tau_h$  is the time constant of inactivation. Steady state voltage-dependent time constants of activation and inactivation functions are as follows:

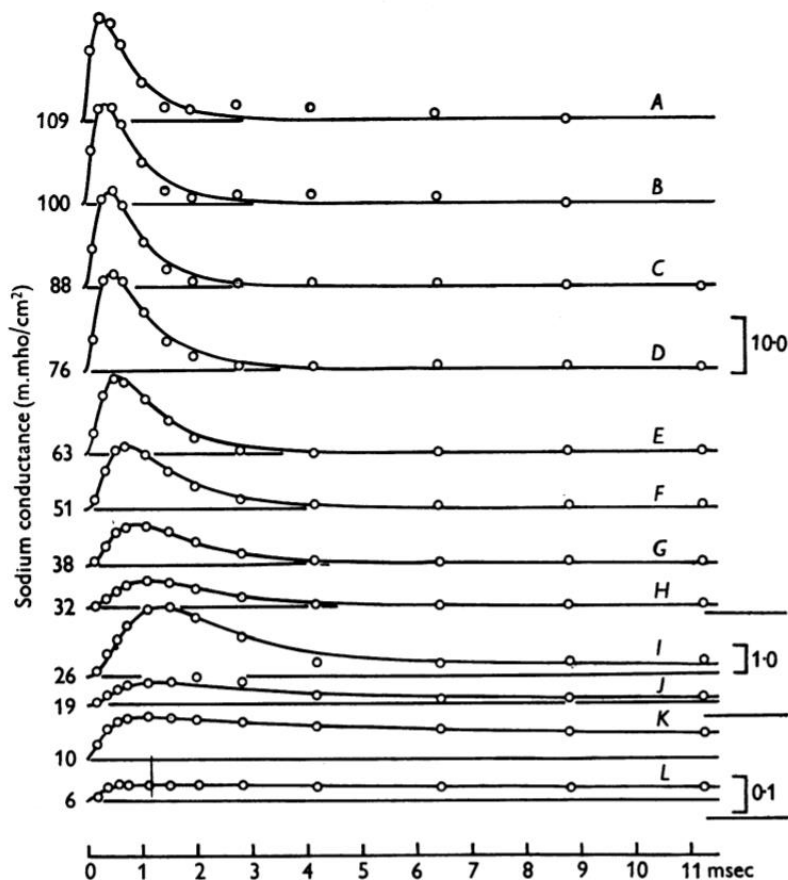
$$\tau_m = \frac{1}{\alpha_m + \beta_m}; \tau_h = \frac{1}{\alpha_h + \beta_h}$$

Ok, so given these steady-state equations, and the following two assumptions: At rest, the sodium conductance is small relative to the conductance during a large depolarization, (1) which therefore allows us (them) to neglect  $m_{\infty}$  if the depolarization is greater than 30 mV. Also, inactivation is "very nearly complete" is the  $V < -30$  mV so that (2)  $h_{\infty}$  may also be neglected.

We can further reduce the equation for sodium conductance to:

$$g_{Na} = \bar{g}_{Na} \left[ 1 - \exp\left(-\frac{t}{\tau_m}\right) \right]^3 \exp\left(-\frac{t}{\tau_h}\right)$$

Then  $\bar{g}_{Na}$ ,  $\tau_m$  and  $\tau_h$  values were calculated by fitting that equation to the following experimental data:



Using these, they were able to calculate  $\alpha$  and  $\beta$  using:  $\alpha_m = m_\infty/\tau_m$  and  $\beta_m = (1 - m_\infty)/\tau_m$  as shown below:

TABLE 2. Analysis of curves in Fig. 6

Curve	V (mV)	$\bar{g}'_{Na}$ (m.mho/cm <sup>2</sup> )	$m_\infty$	$\tau_m$ (msec)	$\alpha_m$ (msec <sup>-1</sup> )	$\beta_m$ (msec <sup>-1</sup> )	$\tau_h$ (msec)	$h_\infty$	$\alpha_h$ (msec <sup>-1</sup> )	$\beta_h$ (msec <sup>-1</sup> )
—	(-∞)	(42.9)	(1.00)	—	—	—	—	—	—	—
A	-109	40.3	0.980	0.140	7.0	(0.14)	0.67	(0)	(0)	1.50
B	-100	42.6	0.997	0.160	6.2	(0.02)	0.67	(0)	(0)	1.50
C	-88	46.8	1.029	0.200	5.15	(-0.14)	0.67	(0)	(0)	1.50
D	-76	39.5	0.975	0.189	5.15	0.13	0.84	(0)	(0)	1.19
E	-63	38.2	0.963	0.252	3.82	0.15	0.84	(0)	(0)	1.19
F	-51	30.7	0.895	0.318	2.82	0.33	1.06	(0)	(0)	0.94
G	-38	20.0	0.778	0.382	2.03	0.58	1.27	(0)	(0)	0.79
H	-32	15.3	0.709	0.520	1.36	0.56	1.33	(0)	(0)	0.75
I	-26	7.90	0.569	0.600	0.95	0.72	(1.50)	(0.029)	(0.02)	(0.65)
J	-19	1.44	0.323	0.400	0.81	1.09	(2.30)	(0.069)	(0.03)	(0.40)
K	-10	0.13	0.145	0.220	0.66	3.9	(5.52)	(0.263)	(0.05)	(0.13)
L	-6	0.046	0.103	0.200	0.51	4.5	(6.73)	(0.388)	(0.06)	(0.09)
—	(0)	(0.0033)	(0.042)	—	—	—	—	(0.608)	—	—

Values enclosed in brackets were not plotted in Figs. 7-10 either because they were too small to be reliable or because they were not independent measurements obtained in this experiment.

$\alpha_h$  and  $\beta_h$  were derived in a similar manner, by plotting against data and solved using:  $\alpha_h = h_\infty/\tau_h$  and  $\beta_h = (1 - h_\infty)/\tau_h$ .

Modeling sodium currents in vestibular ganglion neurons

We used a different but functionally equivalent formulation:

$$I_{Na} = \bar{g}_{Na}(m^3h)(V - E_{Na})$$

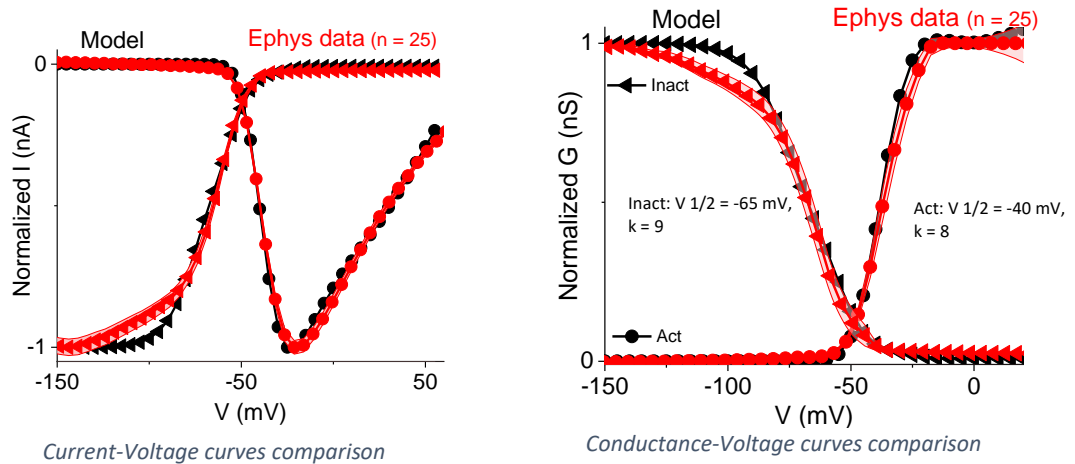
where

$$m' = \frac{m_\infty - m}{\tau_m}; \quad h' = \frac{h_\infty - h}{\tau_h}$$

and

$$m_\infty = \left[ 1 + \exp\left(-\frac{V + V^{1/2}}{k}\right) \right]^{-1}; \quad h_\infty = \left[ 1 + \exp\left(-\frac{V + V^{1/2}}{k}\right) \right]^{-1}$$

$m_\infty$  is still the steady-state activation function,  $\tau_m$  the time constant of activation,  $h_\infty$  the steady-state inactivation function, and  $\tau_h$  the time constant of inactivation. Steady Conductance density ( $\bar{g}$ ), reversal potential ( $E_{Na}$ ), half activation ( $V^{1/2}$ ), and slope factor ( $k$ ) were based on experimentally derived values from this our study.



Our steady state equations are therefore:

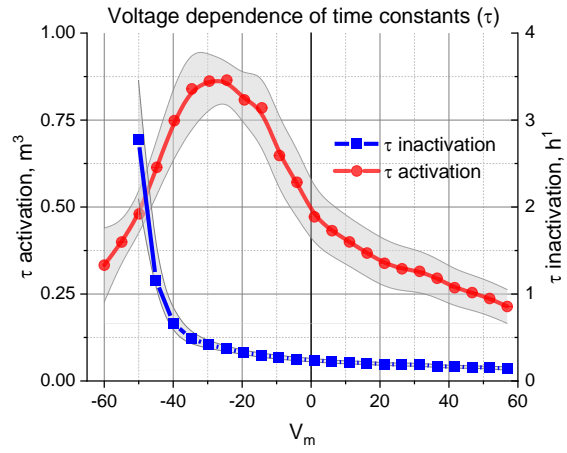
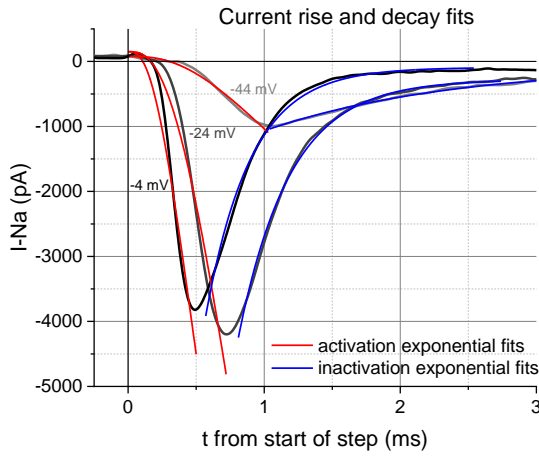
$$m_\infty = \left[ 1 + \exp\left(-\frac{V + 40}{8}\right) \right]^{-1}; \quad h_\infty = \left[ 1 + \exp\left(-\frac{V + 65}{9}\right) \right]^{-1}$$

Our time constants of activation and inactivation were derived by fitting the rising and decay phase of sodium currents to determine the voltage-dependence of time constants. The time constant of activation ( $\tau_m$ ) was assessed by fitting the rising phase of a sodium current with the equation:

$$y = y_0 - A(1 - e^{-\frac{x}{\tau}})^3$$

The power (3) was used since it best fit the very fast rise of the sodium current. The time constant of inactivation ( $\tau_h$ ) was assessed by fitting the decaying phase of a sodium current with:

$$y = y_0 + A(e^{-\frac{x}{\tau}})$$



Using a similar methodology, Rothman and Manis (2003c) use the following equations for  $\tau_m$  and  $\tau_h$ , which were subsequently used in Hight and Kalluri 2016, and Ventura and Kalluri, 2019:

$$\tau_m = 10 \left\{ 5 \exp \left[ \frac{V + 60}{18} \right] + 36 \exp \left[ -\frac{V + 60}{25} \right] \right\}^{-1} + 0.04$$

$$\tau_h = 100 \left\{ 7 \exp \left[ \frac{V + 60}{11} \right] + 10 \exp \left[ -\frac{V + 60}{25} \right] \right\}^{-1} + 0.6$$

where presumably

$$\alpha_m = 5 \exp \left[ \frac{V + 60}{18} \right]; \beta_m = 36 \exp \left[ -\frac{V + 60}{25} \right]$$

$$\alpha_h = 7 \exp \left[ \frac{V + 60}{11} \right]; \beta_h = 10 \exp \left[ -\frac{V + 60}{25} \right]$$

These were putatively derived from data in Costa (1996):

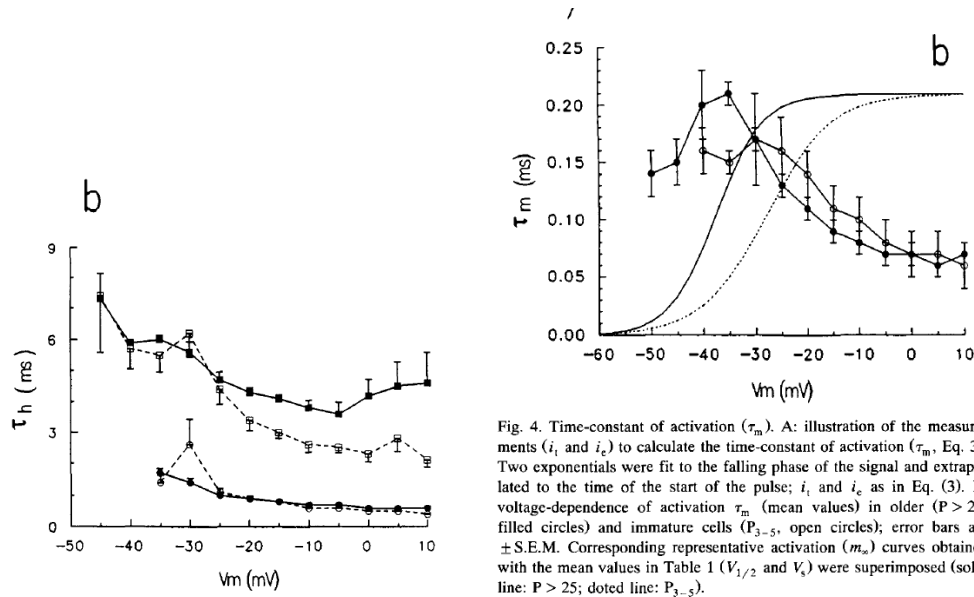


Fig. 4. Time-constant of activation ( $\tau_m$ ). A: illustration of the measurements ( $i_i$  and  $i_e$ ) to calculate the time-constant of activation ( $\tau_m$ , Eq. 3). Two exponentials were fit to the falling phase of the signal and extrapolated to the time of the start of the pulse;  $i_i$  and  $i_e$  as in Eq. (3). B: voltage-dependence of activation  $\tau_m$  (mean values) in older ( $P > 25$ , filled circles) and immature cells ( $P_{3-5}$ , open circles); error bars are  $\pm$  S.E.M. Corresponding representative activation ( $m_\infty$ ) curves obtained with the mean values in Table 1 ( $V_{1/2}$  and  $V_s$ ) were superimposed (solid line:  $P > 25$ ; dotted line:  $P_{3-5}$ ).

Left: voltage-dependence of the time-constants of inactivation measured in activation protocols

Citations:

**Costa PF.** The kinetic parameters of sodium currents in maturing acutely isolated rat hippocampal CA1 neurones. *Developmental Brain Research* 91: 29–40, 1996.

**Hight AE, Kalluri R.** A biophysical model examining the role of low-voltage-activated potassium currents in shaping the responses of vestibular ganglion neurons. *Journal of Neurophysiology* 116: 503–521, 2016.

**Hodgkin AL, Huxley AF.** A quantitative description of membrane current and its application to conduction and excitation in nerve. *The Journal of Physiology* 117: 500–544, 1952.

**Rothman JS, Manis PB.** The Roles Potassium Currents Play in Regulating the Electrical Activity of Ventral Cochlear Nucleus Neurons. *Journal of Neurophysiology* 89: 3097–3113, 2003.

**Ventura CM, Kalluri R.** Enhanced Activation of HCN Channels Reduces Excitability and Spike-Timing Regularity in Maturing Vestibular Afferent Neurons. *J Neurosci* 39: 2860–2876, 2019.